

# Product Information

## CF™ Dye dUTP Conjugates

| Catalog number | Size    | Product      | Molecular weight | Ex/Em (nm) |
|----------------|---------|--------------|------------------|------------|
| 40004-T        | 5 nmol  | CF™405S-dUTP | ~1684            | 404/431    |
| 40004          | 25 nmol |              |                  |            |
| 40008-T        | 5 nmol  | CF™488A-dUTP | ~1429            | 490/515    |
| 40008          | 25 nmol |              |                  |            |
| 40002-T        | 5 nmol  | CF™543-dUTP  | ~1385            | 541/560    |
| 40002          | 25 nmol |              |                  |            |
| 40005-T        | 5 nmol  | CF™568-dUTP  | ~1476            | 562/583    |
| 40005          | 25 nmol |              |                  |            |
| 40006-T        | 5 nmol  | CF™594-dUTP  | ~1491            | 593/614    |
| 40006          | 25 nmol |              |                  |            |
| 40007-T        | 5 nmol  | CF™640R-dUTP | ~1594            | 642/662    |
| 40007          | 25 nmol |              |                  |            |
| 40003-T        | 5 nmol  | CF™680R-dUTP | ~1675            | 680/701    |
| 40003          | 25 nmol |              |                  |            |

### Storage and Handling

Store desiccated at  $\leq -20^{\circ}\text{C}$ . When stored as recommended, product is stable for at least 6 months from date of receipt. For aqueous solutions, prepare single use aliquots and store protected from light at  $-20^{\circ}\text{C}$  for up to 6 months. Avoid freeze-thaw cycles. We recommend preparing a 1 mM stock solution in 10 mM Tris pH 7.4.

### Product Application

CF™ dyes are Biotium's next-generation fluorescent dyes, with combined advantages in brightness, photostability, and water solubility compared to other dyes like Alexa Fluor®, DyLight®, Cy® Dye, and IRDye®. CF dye dUTP can be used for TUNEL staining<sup>1</sup>, or can be used in place of dTTP in standard DNA labeling and synthesis protocols to generate fluorescent dsDNA and oligonucleotide probes.

Note: CF™405S-dUTP may not be suitable for TUNEL staining in tissues due to blue autofluorescence in tissues and lower incorporation efficiency in tissue sections compared to other CF™ dye dUTP conjugates.

Note: for PCR applications, Taq polymerase should be used with dUTP conjugates, because dUTP inhibits archaeal polymerases such as *Pfu* and Vent®.<sup>2,3</sup>

### References

- Gold et al. (1994). Lab Invest. 71 (2):219-25.
- Slupphaug et al. (1993). Anal Biochem. 211 (1):164-9.
- Hogrefe et al. (2002). PNAS 99 (2): 596-601.

## Protocols

### DNA labeling by PCR

#### 1. Materials Required but not Provided

- Taq DNA polymerase (see note under product application)
- 10X Taq reaction buffer
- 25 mM  $\text{MgCl}_2$
- dATP, dTTP, dCTP, dGTP (separate solutions), 1 mM each
- DNA template
- Forward and reverse primers, 10  $\mu\text{M}$  each
- PCR clean-up kit (optional)

#### 2. PCR reaction

2.1 For each labeling reaction, set up the PCR reaction mix as shown below:

| Component                             | Volume per reaction       | Final concentration (after addition of dUTP) |
|---------------------------------------|---------------------------|--|
| 10X Taq reaction buffer               | 2 $\mu\text{L}$           | 1X   |
| 25 mM $\text{MgCl}_2$                 | 2 $\mu\text{L}$           | 5 mM   |
| 1 mM dATP                             | 2 $\mu\text{L}$           | 100 $\mu\text{M}$                            |
| 1 mM dCTP                             | 2 $\mu\text{L}$           | 100 $\mu\text{M}$                            |
| 1 mM dGTP                             | 2 $\mu\text{L}$           | 100 $\mu\text{M}$                            |
| 1 mM dTTP                             | 1 $\mu\text{L}$           | 50 $\mu\text{M}$                             |
| 10 $\mu\text{M}$ forward primer       | 1 $\mu\text{L}$           | 500 nM                                       |
| 10 $\mu\text{M}$ reverse primer       | 1 $\mu\text{L}$           | 500 nM                                       |
| Template                              | 1 ng                      | 50 $\mu\text{g}/\mu\text{L}$                 |
| Taq                                   | 1 U                       | 0.05 U/ $\mu\text{L}$                        |
| Molecular grade $\text{dH}_2\text{O}$ | to 19 $\mu\text{L}$ total |  |

2.2 Add 1  $\mu\text{L}$  of 1 mM CF dye dUTP to the reaction tube.

Optional: for an unlabeled control reaction, add 1  $\mu\text{L}$  of 1 mM dTTP instead of CF dye dUTP.

2.3 Perform PCR according to the following cycling protocol:

|  |           |
|--|-----------|
| Denaturing/hot-start Taq activation<br>94°C, 2 min. (see note 1) | Hold      |
| Denaturing 94°C 30 sec.  | Cycle 30X |
| Annealing (see note 2) 30 sec.                                   |           |
| Extension 72°C 1 min. (see note 3)                               |           |
| Final extension 72°C 5 min.                                      | Hold      |

#### Notes:

1. This protocol was optimized for Cheetah™ Hot Start Taq polymerase (see related products). Other hot-start Taq polymerases may require longer activation times.
2. Set the annealing temperature 5°C below the melting temperature ( $T_m$ ) of your primers.
3. This cycling protocol was optimized for 200-300 bp amplicons. Longer amplicons may require longer extension times.

2.4 Optional: use a PCR clean-up kit to remove unincorporated nucleotides.

2.5 Run 10% of the labeled product on an agarose gel without DNA dye added to analyze the efficiency and specificity of the PCR reaction. CF dye fluorescence can be imaged on a UV light box or laser-based gel scanner. Note: Far-red fluorescence emission (650 nm or longer) is not visible to the human eye.

2.6 Post-stain the gel with DNA gel stain to image the total PCR product or optional unlabeled control PCR product.

## Terminal deoxynucleotidyl transferase dUTP nick end labeling (TUNEL) of apoptotic cells

Note: Biotium also offers CF Dye TUNEL Assay Kits with a selection of dye colors, which include Equilibration Buffer, CF dye TUNEL reaction buffer, and TdT enzyme (see related products).

### 1. Materials Required but not Provided

- Phosphate buffered saline pH 7.4 (PBS)
- 4% formaldehyde/PBS
- 70% ethanol (optional)
- PBS/0.2% Triton™ X-100
- PBS/0.1% Triton™ X-100/5 mg/mL bovine serum albumin (BSA)
- 12.5 U/uL recombinant terminal transferase (TdT) enzyme
- 5X TdT reaction buffer: 1M potassium cacodylate, 125 mM Tris-HCl, 1.25 mg/mL BSA, pH 6.6
- 25 mM CoCl<sub>2</sub> solution
- 100 μM dATP

### 2. Sample preparation

#### 2.1 Preparation of cells or fresh-frozen tissue sections

- Optional: include an extra sample to perform a negative control TUNEL reaction without TdT enzyme.
- Wash cells or sections twice in PBS.
- Fix samples in 4% formaldehyde in PBS for 30 minutes at 4°C.
- Optional: store cells in 70% ethanol at -20°C for up to two weeks
- Wash twice in PBS.
- Permeabilize in 0.2% TX-100 in PBS for 30 minutes at room temperature.
- Wash twice in PBS.

#### 2.2 Preparation of paraffin tissue sections

- Optional: include an extra sample to perform negative control (no TdT enzyme) TUNEL labeling.
- Deparaffinize and rehydrate sections according to standard protocols.
- Wash twice in PBS.
- Permeabilize sections with 20 μg/mL proteinase K in PBS for 30 minutes at room 37°C. Proteinase K incubation time and temperature may require optimization depending on tissue type. Alternatively, microwave antigen retrieval protocols may be used at this step.
- Wash several times in PBS.

### 3. Reaction mix preparation

#### 3.1 Dilute CF dye-dUTP to 10 μM in dH<sub>2</sub>O.

#### 3.2 Prepare 100 μL of TUNEL equilibration buffer per sample:

- 20 μL 5X TdT reaction buffer
- 20 μL 25 mM CoCl<sub>2</sub>
- 60 μL dH<sub>2</sub>O

#### 3.3 Prepare 50 μL of CF dye TUNEL reaction mix for each sample:

#### TUNEL Reaction Mix

| Component               | Volume per reaction | Final concentration |
|-------------------------|---------------------|---------------------|
| 5X TdT reaction buffer  | 10 μL               | 1X                  |
| 25 mM CoCl <sub>2</sub> | 10 μL               | 5 mM                |
| 100 μM dATP             | 2.5 μL              | 5 μM                |
| 10 μM CF dye-dUTP       | 2.5 μL              | 0.5 μM              |
| 12.5 U/uL TdT           | 1 μL                | 12.5 U/reaction     |
| dH <sub>2</sub> O       | 24 μL               |                     |
| Final volume            | 50 μL               |                     |

Optional: prepare a negative control sample without TdT enzyme.

### 4. TUNEL staining

#### 4.1 Incubate samples with 100 μL equilibration buffer for 5 minutes at room temperature.

- For adherent cells or tissue sections, cover sample with a Parafilm® coverslip to spread buffer evenly over the cells or tissue section.

#### 4.2 Remove equilibration buffer and add 50 μL of reaction buffer to each sample.

- For adherent cells or tissue sections, cover sample with a Parafilm® coverslip to spread buffer evenly over the cells or tissue section.

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#### 4.3 Incubate samples for 60 minutes at 37°C, protected from light. Tissue sections may require 2 hour incubation at 37°C.

- For adherent cells or tissue sections, perform incubation in a humid chamber.
- For cells in suspension, perform incubation in a microplate on a rocking platform, or resuspend cells in reaction buffer every 15 minutes by gently flicking tubes.

#### 4.4 Wash samples 3 x 5 minutes in PBS/0.1% Triton X-100/5 mg/mL BSA.

#### 4.5 Counterstain samples if desired. Mount samples in fluorescence mounting medium and coverslip for microscopy, or analyze cells in suspension by flow cytometry. TUNEL-positive cells should show bright nuclear fluorescence. No staining should be observed in the absence of TdT enzyme.

### Related Products

| Catalog No. | Product  |
|-------------|--|
| 30063       | CF™488A TUNEL Assay Apoptosis Detection Kit        |
| 30064       | CF™594 TUNEL Assay Apoptosis Detection Kit         |
| 30074       | CF™640R TUNEL Assay Apoptosis Detection Kit        |
| 40067       | CF™488A-dCTP                                       |
| 40057       | CF™532-dCTP  |
| 40058       | CF™543-dCTP  |
| 40027       | CF™555 dCTP  |
| 40055       | CF™568-dCTP  |
| 40056       | CF™594-dCTP  |
| 40066       | CF™640R-dCTP                                       |
| 40028       | CF™647 dCTP  |
| 40068       | CF™660R-dCTP                                       |
| 40031       | CF™555 ddCTP                                       |
| 40032       | CF640R UTP   |
| 40001       | 5-Tetramethylrhodamine-dUTP                        |
| 40063       | Fluorescein-12-dUTP                                |
| 40059       | DEAC-dUTP  |
| 40029       | Biotin-11-dUTP                                     |
| 40022       | Biotin-16-dUTP                                     |
| 40030       | Biotin-20-dUTP                                     |
| 40035       | Biotin-11-CTP                                      |
| 40036       | Biotin-11-dCTP                                     |
| 40033       | Biotin-11-UTP                                      |
| 40023       | Biotin-16-UTP                                      |
| 40034       | Biotin-20-UTP                                      |
| 40078       | Digoxigenin-dUTP, alkali stable                    |
| 40020       | 5-Aminoallyl-dUTP                                  |
| 40021       | 5-Aminoallyl-UTP                                   |
| 40052       | dNTP Set, 100 mM each                              |
| 29050       | Cheetah™ Hot Start Taq DNA Polymerase              |
| 41003       | GelRed™ Nucleic Acid Gel Stain, 10,000X in water   |
| 41004       | GelGreen™ Nucleic Acid Gel Stain, 10,000X in water |

Please visit our website at [www.biotium.com](http://www.biotium.com) to view our full selection of CF™ dye bioconjugates, including antibodies, antibody labeling kits, phalloidin, Annexin V and α-bungarotoxin, as well as fluorescent reagents and kits for genomics and cell biology research.

CF dye technology is covered by pending US and international patents.

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