

Product Information

CF™ Dye Conjugated Rabbit Anti-HA Tag Antibodies

| Catalog No. | Product Description |
|-------------|---------------------------|
| 20238 | CF488A Rabbit Anti-HA Tag |
| 20239 | CF594 Rabbit Anti-HA Tag |
| 20240 | CF640R Rabbit Anti-HA Tag |
| 20486 | CF647 Rabbit Anti-HA Tag |

Unit size: 50 uL

Concentration: 1 mg/mL in pH ~7.4 PBS containing 50% glycerol, 2 mg/ml bovine serum albumin (IgG-free and protease-free) and 0.05% sodium azide.

Storage and Handling

Product is stable for at least 6 months at -20°C as an undiluted liquid. Storage of the antibody for more than a day at final working dilution is not recommended. Protect from light.

Spectral Properties

$\lambda_{abs} / \lambda_{em}$ maxima in pH 7.4 PBS buffer

| Conjugate | Abs (nm) | Em (nm) |
|-----------|----------|---------|
| CF488A | 490 | 515 |
| CF594 | 593 | 614 |
| CF640R | 642 | 662 |
| CF647 | 650 | 665 |

Product Description

Rabbit anti-HA tag antibodies react with the hemagglutinin (HA) epitope tag (YPYDVPDYA) and are conjugated to Biotium's bright and photostable CF™ dyes.

General Protocols for Using CF™ Dye Conjugated Antibodies

Recommended Dilution Range

Working concentrations of IgG conjugates range from 1-10 ug/mL for most applications. Optimal concentration of the conjugate should be determined empirically.

Immunofluorescence Protocol for Microscopy

There are many methods for immunofluorescence staining. The protocol below is a general guideline for staining cells and should be optimized or modified to obtain the best results for each particular application.

1. Coverslip preparation for adherent cells

- Culture cells on slide chambers or sterile glass coverslips (with poly-L-lysine coating if cells do not adhere well, see below). We recommend 18 x 18 mm square coverslips in 6-well plates or 4-well chamber slides.
- Allow cells to adhere and treat as desired.
- Rinse cells gently with PBS.

2. Coverslip preparation for non-adherent cells

- Coat coverslips with 0.01% poly-L-lysine solution for 10 minutes at room temperature.
- Aspirate the poly-L-lysine solution and allow coverslips to dry completely.
- Centrifuge cells in medium and resuspend in PBS. Transfer cells to coverslips.
- Incubate for 30-60 minutes. Check for adherence by microscope.

3. Fixation and Staining

- Fix with 4% paraformaldehyde/PBS, 15 min.

- Rinse twice with PBS to remove traces of fixative.
- Permeabilize with 0.1 - 0.5% TritonX-100/PBS, 5-10 min.
- Block with blocking agent such as with 5% BSA or normal goat serum in PBS, 30 min.
- Dilute primary antibody in dilution buffer as recommended in the specific product's datasheet. Overlay enough diluted antibody to cover cells on coverslip (150-200 uL is usually sufficient to cover the surface area) or add to each chamber of the chamber slides. Keep slips covered or in a humidified chamber to avoid evaporation.
- Rinse three times with PBS, 5 min each wash.
- Dilute fluorescent secondary antibody in dilution buffer and incubate for 1 hour at room temperature. Working concentrations of IgG conjugates range from 1-10 ug/mL for most applications. We recommend including samples without primary antibody as a control for background staining. Keep slips covered or in a humidified chamber to avoid evaporation.
- Rinse three times with PBS, 5 min each wash.
- Additional staining with fluorescent nuclear stains or phalloidins can be done at this step.
- Invert each coverslip onto a precleaned slide with mounting media, preferably one with an anti-fade preservative. Seal edges with clear polish if desired.
- Store slides in the dark at 4°C.

Staining Protocol for Flow Cytometry

There are many methods for cell staining. The protocol below is a general guideline for flow cytometry and should be optimized or modified for each application.

- Aliquot 1 X 10⁶ cells into 12 X 75 mm polypropylene tubes for flow cytometry.
- For intracellular staining, fix and permeabilize cells according to standard protocols. Biotium also offers a fixation/permeabilization kit for flow cytometry (see related products).
- Add the primary antibody or isotype control at the appropriate dilution to the assay tubes. Incubate according to manufacturer's instructions.
- Rinse cells twice by centrifugation with 2-3 mL incubation buffer.
- Decant supernatant and re-suspend the pellet in remaining volume of wash.
- Add fluorescent secondary antibody and incubate for 20-30 minutes. Working concentrations of IgG conjugates range from 1-10 ug/mL for most applications.
- Rinse cells twice by centrifugation with 2-3 mL incubation buffer. Centrifuge to collect cells after each wash. Decant supernatant.
- Resuspend cells in 0.5 mL of diluent of choice to analyze on flow cytometer. Acquire data using the correct channel.

Tips and Hints:

- No signal or weak fluorescence intensity may suggest the following: (a) insufficient antibody is present for detection, (b) intracellular target was not accessible, (c) excitation sources are not aligned, (d) target protein is not present or expressed at low levels, (e) fluorochrome has faded, and/or (f) primary and secondary antibodies are not compatible.
- High fluorescence intensity may suggest the following: (a) antibody concentration is too high, (b) excess antibody was not washed away efficiently, and/or (c) blocking was inadequate. Increase antibody dilution and washes.

CF™-labeled antibodies can also be used for staining histological sections from paraffin-embedded or frozen tissues.

References

- Donaldson, J.G. Immunofluorescence staining. (2001) Curr Protoc Cell Biol. Chapter 4: Unit 4.3.
- Blose, S.H. and Feramisco, J.R. (1983) Fluorescent methods in the analysis of cell structure. Cold Spring Harbour Laboratory.
- www.chroma.com

Related Products

| Cat.# | Product Name | Unit Size |
|---------|---|-----------|
| 40061-T | RedDot™2 Far Red Nuclear Counterstain, 200X in DMSO, Trial Size (15-20 tests) | 25 uL |
| 23001 | EverBrite™ Mounting Medium | 10 mL |
| 23002 | EverBrite™ Mounting Medium with DAPI | 10 mL |
| 23003 | EverBrite™ Hardset Mounting Medium | 10 mL |
| 23004 | EverBrite™ Hardset Mounting Medium with DAPI | 10 mL |
| 23005 | CoverGrip™ Coverslip Sealant | 15 mL |
| 22005 | Mini Super ^{HT} Pap Pen 2.5 mm tip, ~400 uses | 1 pen |
| 22006 | Super ^{HT} Pap Pen 4 mm tip, ~800 uses | 1 pen |
| 22015 | Fixation Buffer | 100 mL |
| 22016 | Permeabilization Buffer | 100 mL |
| 22017 | Permeabilization and Blocking Buffer | 100 mL |
| 22010 | 10% Fish Gelatin Blocking Buffer | 100 mL |
| 22011 | Fish Gelatin Powder | 2 x 50 g |
| 22014 | 30% Bovine Serum Albumin Solution | 100 mL |
| 22002 | Tween®-20 | 50 mL |
| 22020 | 10X Phosphate-Buffered Saline (PBS) | 4 L |
| 22023 | Paraformaldehyde, 4% in PBS, Ready-to-Use Fixative | 5 x 20 mL |

Please visit www.biotium.com to view our full selection of products featuring bright and photostable fluorescent CF™ dyes, including streptavidin and other bioconjugates, secondary antibodies, Mix-n-Stain™ antibody labeling kits, and R-PE conjugates. Biotium also offers a variety of apoptosis and cell viability assays for flow cytometry analysis, including mitochondrial membrane potential dyes, fluorescent Annexin V conjugates, and NucView™488 Caspase-3 Substrate for live cells.

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