

Product Information

Mix-n-Stain™ Antibody Labeling Kits

Size: 1 labeling per kit

Storage: -20°C

Stability: Stable for at least 3 months from date of receipt when stored as recommended.

Components:

Component	5-20 ug kit	20-50 ug kit	50-100 ug kit
Dye vial	1 vial Component A	1 vial Component A	1 vial Component A
Mix-n-Stain reaction buffer, 10X	1 vial of 15 uL 99951-1	1 vial of 15 uL 99951-1	1 vial of 30 uL 99951
Mix-n-Stain antibody storage buffer	1 vial of 60 uL 99952-2	1 vial of 150 uL 99952-1	1 vial of 300 uL 99952
Ultrafiltration vial (MWCO=10K)	1 each 99956	1 each 99956	1 each 99956

Product Application

Mix-n-Stain™ antibody labeling kits contain everything you need to rapidly label an antibody with Biotium's next-generation CF™ dyes or biotin. The labeling procedure comprises simple mixing of your antibody with the reaction buffer and optimally formulated dye provided, followed by a brief incubation. The Mix-n-Stain dye is no longer reactive at the end of the labeling, so the conjugate is ready for staining without further purification. After labeling, the dye is covalently linked to the antibody with a degree of labeling of approximately 4-6 dye molecules per antibody molecule. Mix-n-Stain labeling is covalent, so Mix-n-Stain-labeled antibodies can be used for multicolor fluorescence staining without transfer of dyes between antibodies.

Mix-n-Stain labeling can tolerate sodium azide, and low levels of glycerol, Tris, and glycine. A microcentrifuge ultrafiltration vial is provided in the kit to rapidly remove incompatible small molecule antibody stabilizers before labeling.

The standard Mix-n-Stain labeling protocol can be performed in the presence of up to four-fold excess of BSA or gelatin to IgG (by ug amount). Simply choose the kit size that corresponds to the amount of IgG you wish to label. A modified protocol is provided for labeling IgG in the presence of excess stabilizer protein or ascites fluid.

In this case, choose the kit size that corresponds to the total amount of protein (IgG + stabilizer, or total protein amount in ascites fluid) in the antibody sample you wish to label. The modified protocol also can be used to label amounts of IgG that fall below the lower range of the kit by adding stabilizer protein to the IgG to bring the total protein amount within the kit range. The modified protocol is not recommended for labeling antibodies in crude antiserum or hybridoma cell culture supernatant due to the low concentration of antibody relative to total protein in these formats.

When performing direct immunofluorescence with a fluorescently-labeled antibody, you may need to use a higher concentration of antibody to achieve similar staining intensity compared to indirect immunofluorescence detection using unlabeled primary plus labeled secondary antibody. In our internal testing, indirect immunofluorescence staining results in about 3-fold signal amplification compared to direct immunofluorescence staining.

Labeled secondary antibodies will still bind to primary antibodies labeled using Mix-n-Stain kits, therefore if multiple primary antibodies from the same species are to be used for multicolor immunofluorescence staining, a secondary antibody cannot be used to distinguish an unlabeled primary antibody from a Mix-n-Stain labeled primary antibody from the same species. Mix-n-Stain labeled antibodies can be used as a tertiary staining antibody after standard immunofluorescence staining with primary and secondary antibodies. Biotium also offers biotin Mix-n-Stain labeling kits, for secondary detection using CF™ dye-labeled streptavidin or CF™ dye-labeled monoclonal mouse anti-biotin.

For more information on Mix-n-Stain™ labeling technology, please see the Frequently Asked Questions section on page 3.

Before you begin

Mix-n-Stain antibody labeling kits are optimized for labeling IgG antibodies. We do not recommend using them to label other proteins, because the degree of labeling may not be optimized. Mix-n-Stain labeling conditions may cause IgM antibodies to denature.

Check the compatibility of your antibody with the antibody compatibility guide below. If your primary antibody is a commercial product, please contact the supplier to obtain the antibody concentration and formulation. Mix-n-Stain labeling cannot be used to label antibodies in crude serum or hybridoma supernatants. Use either a protein A purification procedure or a commercial antibody clean-up kit, such as the Pierce Antibody Clean-Up Kit to purify IgG prior to labeling.

An antibody solution free of stabilizers produces the best labeling results, however, low levels of BSA, gelatin, Tris, glycerol in the antibody solution can be tolerated in the standard Mix-n-Stain labeling protocol. The labeling is not affected by sodium azide. For the standard Mix-n-Stain labeling protocol (Section B), select the kit size that corresponds to the total ug amount of IgG you wish to label.

The modified Mix-n-Stain labeling protocol (Section C) is based on the total amount of protein in the labeling reaction rather than the amount of IgG in the labeling reaction. The modified protocol should be used to label antibodies in the presence of excess stabilizer protein. Antibodies in ascites fluid also can be labeled using the modified protocol, however you must determine the concentration of total protein in the ascites fluid before labeling (estimation of protein concentration by measuring absorbance at 280 nm is sufficient). Select the kit size that is appropriate for the total ug amount of protein in the antibody sample that you wish to label. The modified protocol also can be used to label antibody amounts that fall below the lower limit of the kit range by adding additional protein to the IgG to bring the total protein amount within the kit range.

Antibodies labeled in the presence of low levels of BSA and gelatin may show slightly higher background staining compared to antibody labeled without these stabilizers. If the antibody was labeled in the presence of BSA or gelatin, background staining can be greatly reduced by using blocking and wash solutions containing at least 1% BSA or gelatin, respectively.

To remove non-protein components such as Tris, glycine or glycerol, use the ultrafiltration vial provided in the kit to purify your antibody by following the steps in Section A.

The optimal antibody concentration for labeling is 0.5-1 mg/mL. The ultrafiltration vial can be used to concentrate antibody solutions by following the steps in Section A (note: stabilizer proteins will also be concentrated by the ultrafiltration vial). If no antibody clean-up or concentration is required, proceed to the standard antibody labeling protocol (Section B) or the modified antibody labeling protocol (Section C) as appropriate.

Table 1. Mix-n-Stain™ Antibody Compatibility and Labeling Protocol Selection Guide

Component	Compatibility
Sodium Azide	Compatible
Glycerol	≤ 10%: proceed to standard protocol (Section B) > 10%: perform ultrafiltration (Section A)
Tris	≤ 20 mM: proceed to standard protocol (Section B) > 20 mM: perform ultrafiltration (Section A)
Glycine	Perform ultrafiltration (Section A)
BSA or gelatin	≤ 4X IgG by ug amount: use standard protocol (Section B) > 4X IgG by ug amount: use modified protocol (Section C)
Ascites fluid	Use modified protocol (Section C)
Serum	Not compatible; purify IgG
Hybridoma cell culture supernatant	Not compatible; purify IgG

A. Ultrafiltration Protocol

Important: Before you begin, use Table 1 (Mix-n-Stain™ Antibody Compatibility and Labeling Protocol Selection Guide) to determine whether your antibody requires ultrafiltration before labeling. If necessary, contact the manufacturer of your antibody to find out the concentration of IgG and antibody stabilizers. If your antibody does not require ultrafiltration, proceed to the appropriate labeling protocol indicated in Table 1.

The ultrafiltration column membrane has a molecular weight cut-off of 10,000. Therefore, molecules smaller than 10 kDa will flow through the membrane, and molecules larger than 10 kDa, including IgG antibodies, will be retained on the upper surface of the membrane (Figure 1). Take care not to touch the membrane with pipette tips, which could tear or puncture the membrane, resulting in loss of antibody.

Ultrafiltration Vial Capacities

Maximum Sample Volume: 500 µL

Final Concentrate Volume: 15 µL

Filtrate Receiver Volume: 500 µL

Hold-up Volume (Membrane/Support): < 5 µL

1. Add an appropriate amount of antibody to the membrane of the ultrafiltration vial, being careful not to touch the membrane. Spin the solution at 14,000 x g in a microcentrifuge for one minute. Check to see how much liquid has filtered into the filtrate collection tube (lower chamber). Repeat the centrifugation until all of the liquid has filtered into the collection tube. Discard the liquid in the collection tube.
2. For antibody concentration, proceed to Step 3. For clean-up, add an equal volume of 1X PBS to the membrane. Spin the vial at 14,000 x g until the liquid has filtered into the filtrate receiving tube.
3. Add an appropriate concentration of PBS to the membrane to obtain a final antibody concentration of 0.5 - 1 mg/mL. Carefully pipet the PBS up and down over the upper surface of the membrane to recover and resuspend the antibody.
4. Transfer the recovered antibody solution to a fresh microcentrifuge tube.
5. If you are using the modified antibody labeling protocol, save the ultrafiltration vial to concentrate your antibody after labeling. Additional ultrafiltration vials also can be purchased separately (cat. no. 22004).

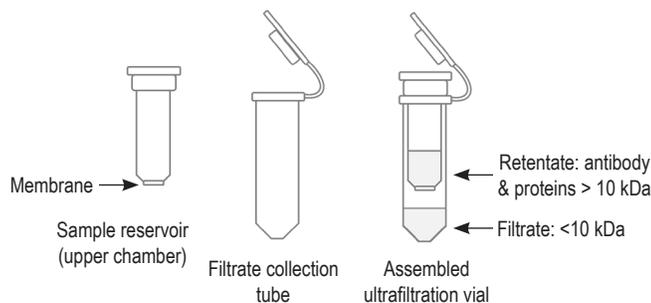


Figure 1. Ultrafiltration vial components.

B. Standard Mix-n-Stain Labeling Protocol

Important: Before you begin, use Table 1 (Mix-n-Stain™ Antibody Compatibility and Labeling Protocol Selection Guide) to determine whether your antibody formulation and concentration are compatible with Mix-n-Stain Labeling, and which labeling protocol you should use. If necessary, contact the manufacturer of your antibody to find out the concentration of IgG and antibody stabilizers.

1. Use your antibody at a concentration of 0.5-1 mg/mL for optimal labeling. If the antibody is in a lyophilized form or is more concentrated, reconstitute or dilute the antibody in PBS. Transfer the antibody to be labeled to a clean tube. Make sure the µg amount of IgG in the labeling reaction falls within the range of the kit.
2. Warm up the Mix-n-Stain™ Reaction Buffer vial and the Mix-n-Stain™ Storage Buffer vial to room temperature before use. Centrifuge the vials briefly to collect the solutions at the bottom of the vials.

3. Mix the 10X Mix-n-Stain™ Reaction Buffer with the antibody solution at a ratio of 1:10 so that the antibody solution contains a final concentration of 1X Reaction Buffer (for example, mix 9 µL of antibody with 1 µL of 10X reaction buffer). Mix the solutions by pipetting up and down a few times.
4. Transfer the entire solution from Step 3 to the vial containing the CF™ dye. There is no need to measure the amount of the dye in the vial. Vortex the vial for a few seconds.
5. Incubate the vial in the dark for 30 minutes at room temperature.
6. Dilute the labeled antibody solution with the provided Storage Buffer. Simply transfer the entire labeled antibody solution into the Storage Buffer. The antibody is now ready to use for staining. The concentration of the CF dye-labeled antibody is approximately the amount of your starting antibody divided by the total volume. **Note:** Antibody Storage Buffer contains 2 mM sodium azide.
7. The labeled antibody is stable for at least 6 months when stored 4°C, protected from light. Alternatively, the antibody can be stored in single use aliquots at -20°C for longer term storage. Note: If you prefer not to use the antibody dilution buffer, you can store the solution in single use aliquots at -20 °C. Without repeated freeze-thaws, the labeled antibody solution should be stable for at least 6 months.

C. Modified Mix-n-Stain Labeling Protocol

Important: Before you begin, use Table 1 (Mix-n-Stain™ Antibody Compatibility and Labeling Protocol Selection Guide) to determine whether your antibody formulation and concentration are compatible with Mix-n-Stain Labeling, and which labeling protocol you should use. If necessary, contact the manufacturer of your antibody to find out the concentration of IgG and antibody stabilizers.

1. Use your antibody solution at a concentration of 0.5-1 mg/mL total protein (IgG plus stabilizer protein) for optimal labeling, using PBS to dilute the solution if necessary. Make sure the µg amount of total protein in the labeling reaction falls within the range of the kit. If you wish to label an amount of IgG that falls below the lower limit of the kit, add BSA to bring to the total protein concentration (IgG + BSA) within the range of the kit and proceed with labeling based on total protein concentration.
2. Warm up the Mix-n-Stain™ Reaction Buffer vial and the Mix-n-Stain™ Storage Buffer vial to room temperature before use. Centrifuge the vials briefly to collect the solutions at the bottom of the vials.
3. Mix the 10X Mix-n-Stain™ Reaction Buffer with the antibody solution at a ratio of 1:10 so that the antibody solution contains a final concentration of 1X Reaction Buffer (for example, mix 9 µL of antibody with 1 µL of 10X reaction buffer). Mix the solutions by pipetting up and down a few times.
4. Transfer the entire solution from Step 3 to the vial containing the CF™ dye. There is no need to measure the amount of the dye in the vial. Vortex the vial for a few seconds.
5. Incubate the vial in the dark for 30 minutes at room temperature.
6. Optional: you can transfer the entire labeling reaction to the tube of antibody storage buffer provided. However, this may result in a highly dilute IgG solution, which may not be practical for subsequent use. To transfer the antibody to storage buffer without additional dilution, follow the steps below. **Note:** Antibody Storage Buffer contains 2 mM sodium azide.
7. Transfer the labeling reaction to the membrane of the ultrafiltration vial provided (or saved after antibody clean-up, above). Centrifuge the vial at 14,000 x g until all of the liquid has filtered into the receiving vial as described in section A.
8. Resuspend the labeled antibody in antibody storage buffer at the desired final concentration of IgG. Carefully pipette the storage buffer up and down over the upper surface of the membrane to recover and resuspend the antibody. **Note:** Antibody Storage Buffer contains 2 mM sodium azide.
9. Transfer the recovered antibody solution to a fresh microcentrifuge tube. The antibody is now ready to use for staining.
10. The labeled antibody is stable for at least 6 months when stored at 4°C, protected from light. Alternatively, the antibody can be stored in single use aliquots at -20°C for longer term storage.

Mix-n-Stain Antibody Labeling Kits Frequently Asked Questions

Question	Answer
How do I remove any unconjugated free dye from the labeled antibody since there is no purification step?	This question relates to a key element of our invention. The unique formulations of our dyes and buffers and the labeling strategy have completely removed this concern, which normally has to be dealt with when using conventional antibody labeling methodology. The exact mechanism on how this problem is solved is proprietary information.
Can I use Mix-n-Stain labeled antibodies for multi-color immunofluorescence staining, or will the dye transfer between antibodies?	Mix-n-Stain labeling results in covalent linkage of dye and antibody, so there will be no dye diffusion or transfer. Please refer to the Product Application section on page 1 for more details.
Can I use a Mix-n-Stain kit for labeling proteins other than antibodies?	Mix-n-Stain kits are optimized for labeling IgG antibodies. We do not recommend them for labeling other proteins. Mix-n-Stain labeling conditions may cause denaturation of IgM antibodies.
Is staining with Mix-n-Stain labeled antibodies as sensitive as staining with unlabeled primary and fluorescent secondary antibodies?	Direct immunofluorescence detection can be less sensitive than indirect detection. Please refer to the Product Application section on page 1 for more details.
What are the advantages of using directly labeled conjugates compared to indirect staining with labeled secondary antibodies?	Direct immunofluorescence staining eliminates the need for secondary antibody incubation and wash steps, and allows the use of multiple primary antibodies from the same species for multicolor detection, or staining of animal tissues with antibodies raised in the same species without secondary antibody cross-reactivity (e.g. mouse-on-mouse staining).
What are the advantages of Mix-n-Stain kits over Invitrogen's Zenon® antibody labeling kits?	The major advantages are: 1) the CF dye is covalently attached to the antibody to eliminate dye transfer or diffusion between antibodies during multi-color staining; 2) Mix-n-Stain conjugates are stable for several months in storage buffer whereas Zenon labeling reagents are required to be used within 30 minutes; 3) Mix-n-Stain conjugates are less bulky because the dyes are directly linked to the antibody, unlike Zenon conjugates which use antibody fragments; 4) No dye/protein optimization is needed, just mix and then stain; 5) No post-staining fixation is required with Mix-n-Stain; 6) Unlike Zenon, Mix-n-Stain labeling is not species-specific.
What are the advantages of Mix-n-Stain kits over Innova Bioscience's Lightning-Link™ Rapid antibody labeling kits?	Mix-n-Stain antibody labeling kits use novel CF dyes which are brighter and more photostable than the dyes provided in Lightning Link kits. Mix-n-Stain kits are sized for labeling smaller amounts of antibody and are sold as single labelings, providing more flexibility compared to Lightning Link kits.
What are CF dyes?	CF dyes are highly water soluble, small organic dyes designed by scientists at Biotium for labeling proteins and nucleic acids. With a series of more than 20 colors, many of our CF dyes are brighter and more photostable than competing dyes. For more detailed comparisons, please see the product flyers for individual CF dyes and the CF dye brochure and FAQ at www.biotium.com .
How do I select a Mix-n-Stain kit?	For each CF dye, there are three labeling kits for labeling of antibody quantities in three different ranges: 1) 5-20 ug, 2) 20-50 ug, and 3) 50-100 ug. For antibody labeling in the absence of stabilizer protein, select a kit that matches with the amount of your antibody. For antibody labeling in the presence of stabilizer protein or ascites fluid, see Table 1 of the product protocol.
If my antibody amount falls between two kits, which one should I use? For example, if I want to label 50 ug of antibody, should I purchase the 50-100 ug kit or the 20-50 ug kit?	Although either kit will produce good results, it is better to use the smaller kit size if your antibody amount falls between two kit sizes.
What dye/protein ratio should I use to ensure optimal labeling?	There is no need to measure the dye amount or vary the reaction time as long as the amount of your antibody to be labeled falls within the range specified for each kit. With Mix-n-Stain labeling kits optimal labeling is ensured because of the proprietary dyes and reaction buffer.
Can I split the kit contents and use it more than one time?	No. The Mix-n-Stain kits are optimized for 1 labeling. We do not recommend that you try to split the kit to label more than one antibody or for more than one use.
How important is the antibody concentration?	The kits are optimized for labeling antibodies with a concentration between 0.5-1.0 mg/mL. If your antibody solution is too dilute, you can concentrate it by centrifugation using the ultra-filtration vial provided in the kit. If your antibody solution is too concentrated, you can dilute it with 1x PBS. Antibody concentrations outside the recommended range may result in either under- or over-labeling.
I performed immunofluorescence staining with my labeled antibody, but I don't see any signal. What should I do?	<p>Check with the antibody manufacturer to confirm that the antibody formulation and concentration are compatible with the kit labeling protocol you selected.</p> <p>You should confirm that your primary antibody is sensitive and specific for your application using indirect labeling before attempting direct labeling. You may need to use a higher concentration of primary antibody to achieve similar signal intensity with direct labeling as with indirect labeling. Please refer to the Product Application section on page 1 for more information.</p> <p>Covalent labeling may affect the reactivity of certain antibodies. You can test if this is the case by performing indirect immunofluorescence labeling with your Mix-n-Stain labeled primary with secondary detection using a fluorescently-labeled secondary antibody to confirm that the primary antibody is still reactive.</p> <p>If you have access to a fluorescence gel reader or scanner that is compatible with the excitation/emission wavelengths of the dye you are using, you can confirm labeling of your antibody by performing denaturing SDS-PAGE on a small amount (0.1-0.5 ug) of labeled antibody, then imaging the gel fluorescence. You should be able to detect fluorescent bands representing IgG heavy and light chains at ~55 kDa and ~25 kDa.</p>

Ordering Information

Mix-n-Stain™ Antibody Labeling Kit			Catalog Number		
Label/Dye	Ex (nm)	Em (nm)	1 X (5-20 ug) labeling	1 X (20-50 ug) labeling	1 X (50-100 ug) labeling
Biotin	N/A	N/A	92244	92266	92286
FITC	494	518	92294	92295	92296
CF™350	347	448	92270	92250	92230
CF™405S	404	431	92271	92251	92231
CF™405M	408	452	92272	92252	92232
CF™488A	490	515	92273	92253	92233
CF™532	527	558	92289	92290	92291
CF™543	541	560	92287	92267	92247
CF™555	555	565	92274	92254	92234
CF™568	562	583	92275	92255	92235
CF™594	593	614	92276	92256	92236
CF™633	630	650	92277	92257	92237
CF™640R	642	662	92278	92258	92245
CF™647	650	665	92279	92259	92238
CF™660C	667	685	92280	92260	92239
CF™660R	663	682	92281	92261	92243
CF™680	681	698	92282	92262	92240
CF™680R	680	701	92283	92263	92246
CF™750	755	777	92284	92264	92241
CF™770	770	797	92285	92265	92242
CF™790	784	806	92288	92268	92248

Related Products

Catalog #	Product Name	Unit Size
22004	Ultrafiltration vial	5 per pack
40061-T	RedDot™2 Far Red Nuclear Counterstain, 200X in DMSO, Trial Size	25 uL (15-20 tests)
23001	EverBrite™ Mounting Medium	10 mL
23002	EverBrite™ Mounting Medium with DAPI	10 mL
23003	EverBrite™ Hardset Mounting Medium	10 mL
23004	EverBrite™ Hardset Mounting Medium with DAPI	10 mL
23005	CoverGrip™ Coverslip Sealant	15 mL
22005	Mini Super ^{HT} Pap Pen 2.5 mm tip, ~400 uses	1 pen
22006	Super ^{HT} Pap Pen 4 mm tip, ~800 uses	1 pen
30069	AccuEasy™ Flow Cytometry Kit	1 kit
23006	Flow Cytometry Fixation/Permeabilization Kit	50 tests
22015	Fixation Buffer	100 mL
22016	Permeabilization Buffer	100 mL
22017	Permeabilization and Blocking Buffer	100 mL
22010	10% Fish Gelatin Blocking Buffer	100 mL
22011	Fish Gelatin Powder	2 x 50 g
22014	30% Bovine Serum Albumin Solution	100 mL
22002	Tween®-20	50 mL

Please visit www.biotium.com to view our full selection of products featuring bright and photostable CF™ dyes, including secondary antibodies, streptavidin, anti-biotin, and anti-tag antibodies. Biotium also offers a variety of apoptosis and cell viability assays for flow cytometry analysis, including mitochondrial membrane potential dyes, fluorescent Annexin V conjugates, and NucView™488 Caspase-3 Substrate for live cells.

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