**GelGreen® Nucleic Acid Gel Stain, 10,000X**

**Catalog Numbers:**
- 41004 (in DMSO)
- 41005 (in water)

**Unit Size:** 0.5 mL (41004 and 41005), 0.1 mL (41005-T), 10 mL (41005-1)

**Storage and Handling**
GelGreen® is a very stable dye. Store 10,000X solution and dilute solutions of GelGreen® at room temperature, protected from light. Dye precipitation may occur at lower temperatures, resulting in lower signal or the appearance of precipitate on the surface of the gel. If this occurs, heat the solution to 45-50°C for two minutes and vortex. GelGreen® is stable for at least one year from the date it is received.

**Product Description**
GelGreen® is a sensitive, stable and environmentally safe green fluorescent nucleic acid dye specifically designed for gel staining. GelGreen® has UV absorption between 250 nm and 300 nm and a strong absorption peak centered around 500 nm (Figure 1). Thus, GelGreen® is compatible with either a 254 nm UV transilluminator or a gel reader equipped with visible light excitation (such as a 488 nm laser-based gel scanner or a Dark Reader). GelGreen® is far more sensitive than SYBR® Safe (Figure 2). Unlike SYBR® dyes, which are known to be unstable, GelGreen® is very stable, both hydrolytically and thermally.

GelGreen® was subjected to a series of tests at Biotium and by three independent testing services to assess the dye’s safety for routine handling and disposal. Test results confirm that the dye is impermeable to both latex gloves and cell membranes (Figure 3). Unlike the highly mutagenic EtBr and the reportedly mutation-enhancing SYBR® Green I (1), GelGreen® is noncytotoxic and nonmutagenic at concentrations well above the working concentrations used in gel staining, because of the dye’s inability to cross cell membranes. GelGreen® successfully passed environmental safety tests in compliance with CCR Title 22 Hazardous Waste Characterization, under which GelGreen® is classified as non-hazardous waste. A complete safety report is available at www.biotium.com.

GelGreen® Nucleic Acid Gel Stain, 10,000X is a concentrated GelGreen® solution that can be diluted 10,000 times for use in precast gel staining or ~3,300 times for use in post gel staining according to the procedures described below. One vial (0.5 mL) of 10,000X solution can be used to prepare at least 100 precast minigels or post-stain at least 100 minigels.

Gel staining with GelGreen® is compatible with downstream applications such as gel extraction and cloning. GelGreen® is efficiently removed from DNA by phenol/chloroform extraction and ethanol precipitation. GelGreen® is not designed for qPCR applications, for which we recommend EvaGreen® dye (cat# 31000).

**Spectral Characteristics**

![Fluorescence](image)

Figure 1. Excitation (left) and emission (right) spectra of GelGreen® bound to dsDNA in TBE buffer.

**Staining Protocols**

Because nucleic acid binding dyes can affect DNA migration during electrophoresis, post-staining of gels is highly recommended. Post-staining with GelGreen® results in superior sensitivity and eliminates the possibility of dye interference with DNA migration. Post-staining with GelGreen® is simple, requiring no destaining and no special buffer. GelGreen® also can be included in agarose gels using the precast method. While the precast protocol is more convenient, some DNA samples may experience migration retardation or compromised resolution in the presence of GelGreen®. Thus, the post-staining and precast protocols should be compared to determine which one better meets your needs.

Although GelGreen® has undergone extensive safety testing, Biotium recommends following universal safety precautions when working in the laboratory.

**Post-staining Protocol**

1. Run gels as usual according to your standard protocol.
2. Dilute the GelGreen® 10,000X stock reagent ~3,300 fold to make a 3X staining solution in H₂O. Note: including 0.1 M NaCl in the staining solution enhances sensitivity, but may promote dye precipitation if the gel stain is reused.
3. Carefully place the gel in a suitable polypropylene container. Gently add a sufficient amount of the 3X staining solution to submerge the gel.
4. Agitate the gel gently at room temperature for ~30 minutes.
5. Image the stained gel with a 254 nm transilluminator, a Dark Reader® or a similar transilluminator, or a laser-based gel scanner using a long path green filter such as a SYBR® filter or GelStar® filter.
6. Staining solution can be reused at least 2-3 times. Store staining solution at room temperature protected from light.

**Precast Protocol**

Note: The precast protocol is not recommended for polyacrylamide gels. Use the post staining protocol for acrylamide gels.

1. Prepare molten agarose gel solution using your standard protocol.
2. Dilute the GelGreen® 10,000X stock reagent into the molten agarose gel solution at 1:10,000 and mix thoroughly. GelGreen® can be added while the solution is still hot.
3. Cast the gel and allow it to solidify. Load samples and run the gels using your standard protocol.
4. Image the stained gel with a 254 nm transilluminator, a Dark Reader® or a similar transilluminator, or a laser-based gel scanner using a long path green filter such as a SYBR® filter or GelStar® filter.

**Storing GelGreen® gels**

Leftover gel solution with GelGreen® may be stored at room temperature, protected from light, and re-heated later for additional gel casting. GelGreen® precast gels may be stored for later use for up to a month at room temperature in the dark. Storing GelGreen® precast gels at 4°C can result in dye precipitation and poor performance.

**References**

Question

Can GelGreen® be used to stain ssDNA or RNA?

GelGreen® can be used to stain ssDNA and RNA, but we recommend GelRed® for this application because it is five times more sensitive for single stranded nucleic acids than GelGreen®.

Is GelGreen® compatible with downstream applications such as cloning, ligation and sequencing?

Yes. Biotium’s DNA Gel Extraction Kit (see Related Products), DNA gel extraction kits from Qiagen or Zymo, or phenol-chloroform extraction can be used to remove the dye from DNA.

Is GelGreen® compatible with Southern or northern blotting?

GelGreen® has not been validated in blotting applications.

Can I reuse a GelGreen® precast gel after electrophoresis?

We do not recommend reusing GelGreen® precast gels as signal decreases with subsequent electrophoresis.

How should I dispose of GelGreen®?

GelGreen® has passed the EPA regulated Title 22 test. Some facilities have approved the disposal of GelGreen® directly down the drain. However, because regulations vary, please contact your safety office for local disposal guidelines. GelGreen® can be adsorbed to activated charcoal (see Related Products) for disposal as chemical waste.

What is the lower detection limit of GelGreen®?

Some users have reported being able to detect less than 0.1 ng DNA. However, the limit of detection will depend on instrument capability and exposure settings.

Does GelGreen® need to be used in the dark?

You can use the dye in room light, however we recommend storing the dye in the dark.

Is there a difference between 10,000X GelGreen® in DMSO and water?

The GelGreen® stock in water is a newer and improved product compared to the stock in DMSO. We recommend using GelGreen® in water to avoid the potential hazards of handling DMSO, a solvent that can be absorbed through the skin. We continue to offer GelGreen® in DMSO because some users do not wish to alter their established laboratory protocols.