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Product Information

AccuBlue®™ High Sensitivity dsDNA Quantitation Kits

Catalog Numbers

31006-T, AccuBlue® High Sensitivity dsDNA Quantitation Kit with DNA Standard, Trial Size

31006 AccuBlue®™ High Sensitivity dsDNA Quantitation Kits with 8 DNA Standards

Kit Contents

Component	31006-T 200 assays	31006 1000 assays
AccuBlue® High Sensitivity dsDNA Quantitation Solution	50 mL 31008-TA	250 mL 99940
AccuBlue® High Sensitivity Enhancer (100X)	1 mL 99942	3 X 1 mL 99942
AccuBlue® dsDNA Standard in 10 mM Tris pH 7.5, 1 mM EDTA, 2 mM sodium azide, 10 ng/uL dsDNA from calf thymus	0.5 mL 31006C-T	N/A
AccuBlue® dsDNA Standards in 10 mM Tris pH 7.5, 1 mM EDTA, 2 mM sodium azide, 0, 0.5, 1, 2, 4, 6, 8 and 10 ng/uL dsDNA from calf thymus	N/A	Set of eight 0.5 mL each 31006C

Storage and Handling

Store $k\bar{t}$ at 4°C. Protect quantitation solution from light. The $k\bar{t}$ is stable for at least 12 months from date of receipt when stored as recommended.

Spectral Properties

Best linearity is achieved at Ex/Em 485/530 nm. See Figure 1 for spectra.

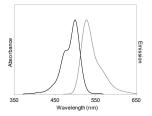
Product Description

The AccuBlue®™ High Sensitivity dsDNA Quantitation Kit with 8 DNA Standards provides ease and simplicity for DNA quantitation. The kit contains AccuBlue® dsDNA Quantitation Solution, Enhancer and pre-diluted dsDNA standards. AccuBlue® assays are based on binding of fluorescent DNA dyes that selectively detect double-stranded DNA over RNA or single-stranded DNA (Figure 2), unlike absorbance-based measurements. The assay is highly reliable in detecting dsDNA ranging from 0.2 to 100 ng (Figure 3), and offers advantages in stability, linear dynamic range, and sensitivity over other traditional methods of DNA quantitation. The assay is tolerant of common contaminants such as proteins, salts, organic solvents and detergents (Table 1). In addition, the AccuBlue® High Sensitivity DNA dye does not readily enter cells, and is non-toxic and non-mutagenic.

The AccuBlue® High Sensitivity dsDNA Quantitation Kit is used with fluorescence 96-well plate readers equipped with fluorescein excitation and emission filters. The assay also can be used with fluorometers such as the Qubit® (Invitrogen) and QuantiFluor™-P (Promega).

AccuBlue® High Sensitivity dsDNA Quantitation Solution is available without standards for users who wish to prepare their own DNA standards. Biotium also offers AccuBlue® Broad Range dsDNA quantitation assays for measuring DNA in the range of 2-2000 ng, and the AccuClear® Ultra High Sensitivity dsDNA quantitation assays for measuring DNA in the range of 0.03-250 ng. See related products for details.

Figure 1: Excitation and emission spectra for AccuBlue® High Sensitivity dsDNA dye in the presence of dsDNA.



Assay Protocol

- Use properly calibrated pipettes and DNase-free pipette tips, tubes and
 plates for best accuracy. It is recommended to test each DNA standard and
 each unknown sample in triplicate. If more than one 96 well plate is to be
 tested in a single assay, it is recommended to include a standard curve on
 each plate to minimize variability between plates.
- 2. Allow the kit components to warm to room temperature before use. Invert the quantitation solution bottle several times and vortex the 100X Enhancer. If precipitation is seen in the enhancer, warm up the vial in a water bath and vortex until dissolved. Before removing the required volume, mix each component well by shaking or vortexing, and centrifuge vials briefly before opening to minimize reagent loss on the cap.
- 3. For 31006, use the set of eight DNA standards provided (31006C). For 31006-T (trial size kit) prepare a set of DNA standards by diluting the 10 ng/uL standard provided in TE buffer (10 mM Tris pH 7-8, 1 mM EDTA) as shown in Table 1 below. The DNA standards can be stored at 4°C for at least 6 months if sodium azide is added to the TE buffer used for dilution at a final concentration of 2 mM.

Table 1. Preparation of DNA standards for 31006-T (trial size kit)

Standard	10 ng/uL DNA standard	TE
10 ng/uL	100 uL	None
8 ng/uL	80 uL	20 uL
6 ng/uL	60 uL	40 uL
4 ng/uL	40 uL	60 uL
2 ng/uL	20 uL	80 uL
1 ng/uL	10 uL	90 uL
0.5 ng/uL	5 uL	95 uL
0 ng/uL	None	100 uL

- Prepare 200uL working solution for each sample to be tested. For one 96well plate, add 200 uL of 100X Enhancer to 20 mL of Quantitation Solution to prepare the working solution, scaling volumes as needed. Mix well. Working solution should be used within an hour of preparation.
- 5. For each sample to be tested, pipette 200 uL of the working solution per well of a black 96-well microplate. To test samples in triplicate, prepare three separate wells for each DNA standard and three separate wells for each unknown DNA sample. Accurate multi-channel pipettes and reagent reservoirs can be used to increase throughput. Black plates are recommended to minimize fluorescence bleed-through between wells. We have found that black 96-well plates from Greiner Bio One or Corning give the most consistent signal-to-noise ratio at low DNA concentrations.
- Add 10 uL of each dsDNA standard and unknown into its own separate well containing working solution and mix well by pipetting up and down.
- Incubate the microplate at room temperature for 5-10 minutes in the dark.
 The assay plate should be read immediately for best results, but can be
 stored and read up to 6 hours after preparation while maintaining accuracy
 from 5-100 ng DNA in the assay.
- Measure fluorescence using a microplate reader to set to 485 nm excitation/530 nm emission maxima or other filter combination for detecting green fluorescence (e.g., FITC filter set).
- Generate a standard curve to determine the unknown DNA concentration (see Figure 2). Average the triplicate values for each sample and subtract

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the average zero DNA value from each data point. Plot the fluorescence values for the DNA standards on the y-axis and ng/well DNA on the x-axis, and fit a trend line through these points to generate a standard curve with a y-intercept = 0. Use the equation for the standard curve trend line to calculate the amount of unknown DNA in each well (y = fluorescence and x = ng DNA per well). Note: the standard curve shown in Figure 2 is for reference only. You must generate your own standard curve using your instrument to calculate the amount of DNA in your unknown samples.

Considerations for Data Analysis

Calf thymus DNA can often serve as a reference for most plant and animal DNA because it is double-stranded, highly polymerized and is approximately 58% AT (42% GC). At times it is preferable to use a dsDNA standard similar to the unknown samples (i.e. similar in size, linear vs. circular). We have found that most linear dsDNA yield similar results; however, it is best to compare the concentration of the unknown sample to a more appropriate standard if necessary. If the fluorescence of an unknown sample is higher than the linear range, further dilute the sample and add 10 uL of the diluted sample to perform the assay. For consistency, it is best to use the same volume in all the wells with samples that do not have high levels of contaminating substances.

Fluorescence quantitation by the AccuBlue® High Sensitivity reagent is linear from 0.2 – 100 ng dsDNA. The dynamic range can be extended to 200 ng with some loss of linearity. If lower end standards are desired, you can further dilute any of the standards with 1X TE to 0.02 ng/uL. Use 10 uL/well to obtain a 0.2 ng/well standard.

Due to differences in instruments, check instrument settings to optimize for the best linearity. Some factors that can affect the final linearity and relative fluorescence intensity are: (1) the excitation and emission wavelengths and bandwidths, (2) cut-off filters, (3) sensitivity settings, (4) pipette accuracy, and (5) microplate manufacturers.

The effects of common DNA contaminants such as salts, solvents, detergents and protein on the AccuBlue® High Sensitivity assay are listed in Table 2. Please also see our AccuClear® Ultra High Sensitivity dsDNA Quantitation Assays (related products), which have different tolerances for certain contaminants compared to AccuBlue® High Sensitivity.

Table 2. Effect of common DNA contaminants on AccuBlue® assay signal

Compound	Initial concentration in DNA sample	Final concentration in assay (200 uL)	Result
Ammonium Acetate	100 mM	5 mM	Pass
Sodium Acetate	600 mM	30 mM	Pass
Sodium Chloride	200 mM	10 mM	Pass
Magnesium Chloride	25 mM	1.25 mM	Pass
Phenol	2 %	0.1 %	Pass
Ethanol	10 %	0.5 %	Pass
Chloroform	2 %	0.1 %	Pass
Sodium Dodecyl Sulfate	0.2 %	0.01 %	Pass
Triton X-100	0.2 %	0.01 %	Pass
CTAB	0.01%	0.0005%	Fail*
Bovine Serum Albumin	200 mg/mL	10 mg/mL	Pass**
dNTPs	2 mM	100 uM	Pass
Polyethylene Glycol	40%	2 %	Pass
Agarose	2%	0.1 %	Pass

^{*}CTAB contamination results in almost complete loss of fluorescence.

Triplicate DNA standard curves were assayed in the presence or absence of the contaminants at the indicated final concentrations. Pass indicates that there was < 20% change in signal in the absence of the contaminant.

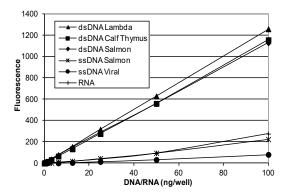


Figure 2: AccuBlue® High Sensitivity dsDNA Quantitation kit selectivity and sensitivity for dsDNA. Triplicate samples of dsDNA or ssDNA from various sources or mouse liver RNA were assayed using AccuBlue® and read at 485/530nm.

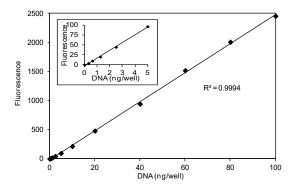


Figure 3: Example of AccuBlue® High Sensitivity dsDNA standard curve.

Related Products

Catalog number	Product
31066-T	AccuGreen™ High Sensitivity dsDNA Quantitation Kit, trial size (for Qubit)
31060-T	AccuBlue®™ NextGen dsDNA Quantitation Kit, trial size
31028	AccuClear® Ultra High Sensitivity dsDNA Quantitation Kit with 7 DNA Standards
31007	AccuBlue®™ Broad Range dsDNA Quantitation Kit with 9 DNA Standards
41003	GelRed™ Nucleic Acid Gel Stain, 10,000X in water
31041-T	Forget-Me-Not® qPCR Master Mix, trial size
31043-T	Forget-Me-Not® Universal Probe Master Mix, trial size

Please visit our website at www.biotium.com for information on our life science research products, including environmentally friendly EvaGreen® qPCR master mixes, fluorescent CF® dye antibody conjugates and reactive dyes, apoptosis reagents, fluorescent probes, and kits for cell biology research.

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^{**}Pass with some perturbation of standard curve linearity.