

# Product Information

## Mix-n-Stain™ Maxi Antibody Labeling Kits

| Catalog No. | CF® Dye     | Ex/Em      |
|-------------|-------------|------------|
| 92420       | CF@350      | 347/448 nm |
| 92421       | CF@405S     | 404/431 nm |
| 92404       | CF@405M     | 408/452 nm |
| 92454       | CF@405L     | 395/545 nm |
| 92405       | CF@488A     | 490/515 nm |
| 92406       | CF@555      | 555/565 nm |
| 92407       | CF@568      | 562/583 nm |
| 92408       | CF@594      | 593/614 nm |
| 92409       | CF@633      | 630/650 nm |
| 92410       | CF@647      | 650/665 nm |
| 92422       | CF@680      | 681/698 nm |
| 92423       | CF@750      | 755/777 nm |
| 92424       | CF@770      | 770/797 nm |
| 92411       | FITC        | 494/518 nm |
| 92415       | Cyanine 555 | 555/565 nm |
| 92419       | Cyanine 647 | 650/665 nm |

### Kit Contents

| Component                                    | Size                       |
|--|----------------------------|
| Dye (Component A)                            | 1 vial (lyophilized solid) |
| Mix-n-Stain™ Reaction Buffer<br>99951-150uL  | 150 uL                     |
| Mix-n-Stain™ Quenching Buffer<br>99854-150uL | 150 uL                     |

**Unit Size:** Single labeling reaction of 1 mg antibody per kit

### Storage and Handling

Store kit at -20°C. Product is stable for at least 12 months from date of receipt when stored as recommended.

### Product Description

Mix-n-Stain™ Maxi Antibody Labeling Kits contain everything you need to rapidly label 1 mg of IgG antibody with Biotium's next-generation CF® Dyes or other fluorescent dyes. Simply mix your antibody with the Reaction Buffer and Dye provided, followed by a 30 minute incubation. After termination of the labeling reaction with the provided Quenching Buffer, the conjugate is ready to use (see Protocol Overview, next page). Any remaining free dye is no longer reactive at the end of the labeling, so the conjugate can be used for staining without further purification. Mix-n-Stain™ labeling is covalent, so labeled antibodies can be used for multiplex staining without transfer of dyes between antibodies.

Mix-n-Stain™ Maxi reactions are optimized for conjugation of 1 mg of IgG. After the reaction, the antibody will be labeled with an average of 4-6 dye molecules per antibody molecule. The kits can be used to label antibody fragments or other proteins, however, the degree of labeling (DOL, or number of dye molecules per antibody molecule) may not be optimal. The kits are not recommended for labeling IgM antibodies. See FAQs (page 3) for information on labeling nanobodies or other proteins.

These kits are not designed to be split for multiple labelings. For smaller labeling reactions, see our Mix-n-Stain™ CF® Dye Antibody Labeling Kits for labeling 5-20 ug, 20-50 ug, or 50-100 ug of antibody. Biotium also offers 3X(5-20 ug) sized kits for multiple labelings of 5-20 ug antibody.

We also offer Mix-n-Stain™ Antibody Labeling Kits for conjugating antibodies to fluorescent proteins (R-PE, APC, PerCP, and tandem dyes) or enzymes (HRP, AP, and glucose oxidase). Our CF® Dye SE Protein Labeling Kits are designed for performing conjugation with post-labeling purification. Visit [www.biotium.com](http://www.biotium.com) for more information.

See Frequently Asked Questions (FAQs) on page 3 for more information about Mix-n-Stain™ labeling technology.

### Kit Compatibility and Considerations for Labeling

Antibodies should be at 1 mg/mL in PBS or similar buffer with no stabilizer proteins, glycerol, Tris, or amino acids such as glycine (See Table 1). Sodium azide does not affect the labeling.

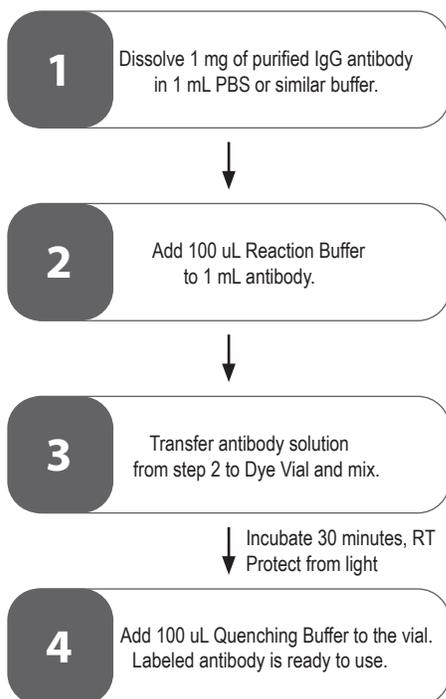
If your antibody concentration is higher than 1 mg/mL, add PBS to dilute it to 1 mg/mL. If your antibody concentration is lower than 1 mg/mL, concentrate the antibody to 1 mg/mL; a 10K MWCO ultrafiltration vial (see Related Products) can be used to concentrate your antibody. Ultrafiltration also can be used to remove incompatible small molecules such as glycerol, Tris, or glycine.

Antibodies containing protein stabilizers such as BSA or gelatin, or crude antibodies in serum, culture supernatant, or ascites fluid should be purified before labeling using a standard Protein A/G or similar purification protocol (ultrafiltration will not remove protein stabilizers).

**Table 1. Reaction Compatibility Guide**

| Component      | Compatibility  |
|----------------|--|
| Sodium Azide   | Compatible   |
| Glycerol       | ≤ 10%: Compatible<br>> 10%: Perform ultrafiltration (see Related Products)     |
| Tris           | ≤ 20 mM: Compatible<br>> 20 mM: Perform ultrafiltration (see Related Products) |
| Glycine        | Perform ultrafiltration (see Related Products)                                 |
| BSA or gelatin | Not compatible; purify antibody  |

### Mix-n-Stain™ Maxi Protocol Overview



### Related Products

| Catalog number | Product   |
|----------------|---|
| 92500-92515    | Mix-n-Stain™ Nanobody Labeling Kits                         |
| 92437          | Mix-n-Stain™ Maxi HRP Antibody Labeling Kit                 |
| 22004          | Ultrafiltration vial, 10K MWCO (pack of 5)                  |
| 22018          | Ultrafiltration vial, 3K MWCO (pack of 5)                   |
| 22011          | Fish Gelatin Powder   |
| 22013          | Bovine Serum Albumin, Fraction V                            |
| 22014          | BSA, 30% Solution   |
| 30071          | AccuOrange™ Protein Quantitation Kit                        |
| 23012          | TrueBlack® IF Background Suppressor System (Permeabilizing) |
| 23013          | TrueBlack® WB Blocking Buffer Kit                           |
| 23007          | TrueBlack® Lipofuscin Autofluorescence Quencher             |
| 23014          | TrueBlack® Plus Lipofuscin Autofluorescence Quencher        |
| 40043          | DAPI in H <sub>2</sub> O, 10 mg/mL                          |
| 40083          | NucSpot® 470 Green Nuclear Counterstain                     |
| 40061          | RedDot™2 Far Red Nuclear Counterstain                       |
| 23008          | Drop-n-Stain EverBrite™ Mounting Medium                     |
| 23009          | Drop-n-Stain EverBrite™ Mounting Medium with DAPI           |
| 23005          | CoverGrip™ Coverslip Sealant                                |
| 22005          | Mini Super <sup>HT</sup> Pap Pen 2.5 mm tip, ~400 uses      |
| 22006          | Super <sup>HT</sup> Pap Pen 4 mm tip, ~800 uses             |
| 23006          | Flow Cytometry Fixation/Permeabilization Kit                |
| 22023          | Paraformaldehyde, 4% in PBS, Ready-to-Use Fixative          |
| 22016          | Permeabilization Buffer                                     |
| 22017          | Permeabilization and Blocking Buffer                        |
| 22030          | AntiFix™ Universal Antigen Retrieval Buffer, 10X            |

Please visit [www.biotium.com](http://www.biotium.com) to view our full selection of products featuring bright and photostable CF® Dyes, including Mix-n-Stain™ Small Ligand Labeling Kits, primary & secondary antibodies, streptavidin, phalloidin, and much more.

### Labeling Protocol

Important: Before you begin, see Kit Compatibility and Considerations for Labeling and Table 1 to verify that your antibody is compatible with Mix-n-Stain™ Maxi labeling.

1. Prepare antibody for labeling at 1 mg/mL. If the antibody is in a lyophilized form or is more concentrated, reconstitute or dilute the antibody in PBS. Transfer 1 mL (1 mg) of antibody to a clean tube.
2. Warm up the Mix-n-Stain™ Reaction Buffer and the Mix-n-Stain™ Quenching Buffer to room temperature before use. Centrifuge the vials briefly to collect the solutions at the bottom of the vials.
3. Add 100 uL of Mix-n-Stain™ Reaction Buffer to 1 mL antibody and mix well.
4. Transfer the entire antibody solution from Step 3 to the vial containing the dye (component A). There is no need to measure the amount of the dye in the vial. Vortex the vial for a few seconds to dissolve the dye.
5. Incubate the vial in the dark for 30 minutes at room temperature with gentle rocking.
6. To terminate the labeling reaction, add 100 uL of Quenching Buffer to the vial containing antibody and dye and mix well. The labeled antibody is now ready to use.

### Storage of Labeled Antibody

The labeled antibody can be stored at 4°C, protected from light, for up to 3 days. For longer-term storage of labeled antibody, we recommend adding stabilizers to the antibody solution, such as sodium azide, BSA (see Related Products), and/or glycerol. Commonly used final concentrations of antibody stabilizers are 0.05% sodium azide, 2 mg/mL BSA or gelatin, and 50% glycerol. Antibodies with BSA and azide can be stored at 4°C, protected from light. Antibodies with BSA and 50% glycerol can be stored at -20°C, protected from light; the glycerol will prevent the solution from freezing, so it is not necessary to aliquot the antibody for storage.

Materials from Biotium are sold for research use only, and are not intended for food, drug, household, or cosmetic use.

CF Dyes and Mix-n-Stain labeling are covered by pending US and international patents.

## Frequently Asked Questions (FAQs) & Troubleshooting

| Question  | Answer  |
|---|---|
| Can Mix-n-Stain™ Antibody Labeling Kits be used to label nanobodies?  | <p>We offer Mix-n-Stain™ Nanobody Labeling Kits, specifically designed for optimized labeling of single-domain nanobodies with CF® Dyes.</p> <p>We also have had customers report successful labeling of nanobodies with our original CF® Dye Mix-n-Stain™ Antibody Labeling Kits. Mix-n-Stain™ kits are optimized for labeling whole IgG (150 kDa) with 4-6 dyes per antibody. Due to their smaller size (~15 kDa), single chain antibodies have fewer potential labeling sites. So to increase the degree of labeling for single chain antibodies, we recommend using a higher ratio of dye to protein. For example, the Mix-n-Stain™ 50-100 ug size kit could be used to label 25-30 ug of nanobody. Also, we do not recommend using the 10K MWCO ultrafiltration vial with proteins smaller than 30 kDa. If ultrafiltration is required prior to Mix-n-Stain™ labeling, a 3K MWCO ultrafiltration vial (Cat. no. 22018) should be used instead.</p> |
| How do I remove any unconjugated free dye from the labeled antibody since there is no purification step?                            | <p>This question relates to a key element of our invention. The unique formulations of our dyes and buffers and the labeling strategy have completely removed this concern, which normally has to be dealt with when using conventional antibody labeling methodology. The exact mechanism on how this problem is solved is proprietary information.</p>  |
| How do I determine the degree of labeling (DOL) of my antibody after the reaction?  | <p>Mix-n-Stain™ kits are formulated to yield DOL of 4-6 dye molecules per antibody molecule. For ease-of-use and convenience, purification is not necessary before using the labeled antibody for staining. If you wish to measure DOL, you can use ultrafiltration (see Related Products) to purify the conjugate, then resuspend in PBS with no stabilizer proteins.</p> <p>For performing reactions to optimize DOL for your antibody or application, we recommend using our CF® Dye SE Protein Labeling Kits. The kits include everything you need to perform controlled conjugation reactions and post-labeling purification, as well as detailed instructions and correction factors for determining DOL for CF® Dyes.</p>  |
| Can I use Mix-n-Stain™ labeled antibodies for multi-color immunofluorescence staining, or will the dye transfer between antibodies? | <p>Mix-n-Stain™ labeling results in covalent linkage of dye and antibody, so there will be no dye diffusion or transfer.</p>  |
| Can I use a Mix-n-Stain™ kit for labeling proteins other than antibodies?   | <p>Mix-n-Stain™ kits are optimized for labeling IgG antibodies, but can be used to label other proteins. Customers have reported successful labeling of nanobodies and single chain antibodies. There are also published reports of Mix-n-Stain™ labeling of enzymes and lectins. However, Mix-n-Stain™ labeling conditions may cause denaturation of IgM antibodies. Note that any conjugation method, including Mix-n-Stain™, may affect the biological activity of proteins. Also, some free unreactive dye may remain after Mix-n-Stain™ labeling, which could interfere with live cell staining or trafficking studies using fluorescently labeled proteins. Ultrafiltration vials can be used to remove free dye after labeling if necessary (see Related Products).</p>  |
| What are the advantages of Mix-n-Stain™ kits over Expedeon Lightning-Link® Rapid antibody labeling kits?                            | <p>Mix-n-Stain™ Antibody Labeling Kits use novel CF® Dyes which are brighter and more photostable than the dyes in Lightning Link® kits. Mix-n-Stain™ kits are sold as single reactions for multiple labeling scales between 5 ug and 1 mg, for greater flexibility.</p>  |
| What are CF® Dyes?  | <p>CF® Dyes are highly water soluble, small organic dyes for labeling proteins and nucleic acids. CF® Dyes are designed to be brighter and more photostable than competing dyes. For more details, visit <a href="http://www.biotium.com">www.biotium.com</a>.</p>  |
| How do I select a Mix-n-Stain™ kit?   | <p>For each CF® Dye, there are three labeling kits for labeling of antibody quantities in different ranges: 5-20 ug, 20-50 ug, 50-100 ug, or 1 mg. Select a kit that matches the amount of your antibody. Mix-n-Stain CF® Dye kits for less than 1 mg antibody can be used for labeling antibody in the presence of excess stabilizer protein or ascites fluid using a modified protocol based on the total amount of protein present. See the relevant kit product information sheet for more information.</p>   |
| What dye/protein ratio should I use to ensure optimal labeling?   | <p>There is no need to measure the dye amount or vary the reaction time as long as the amount of your antibody to be labeled falls within the range specified for the kit.</p>  |
| Can I split the kit contents and use it more than one time?   | <p>No. The Mix-n-Stain™ kits are optimized for a single labeling reaction. We do not recommend that you try to split the kit to label more than one antibody or for more than one use.</p>  |
| How important is the antibody concentration in the labeling reaction?   | <p>Using a higher or lower antibody concentration may result in either under- or over-labeling. We recommend using antibody at 1 mg/mL for Mix-n-Stain™ Maxi labeling.</p>  |

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