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Product Information

RNaseReveal™ Activity Assay Kits

Kit Contents

Kit	Cat.	Component 1:	Component 2:
	No.	Substrate	Assay Buffer
RNaseReveal™	31086	31086A	31086B
Activity Assay Kit		1 mL	1 mL
RNaseReveal™ Red	31087	31087A	31086B
Activity Assay Kit		1 mL	1 mL

Unit Size: 100 assays

Storage and Handling

Store at -20°C. Protect RNaseReveal™ Substrates from light. Kit components are stable for at least 6 months from date of receipt when stored as recommended.

RNaseReveal™ Substrates are RNA-based substrates and precautions should be taken to avoid RNase contamination.

Product Description

The RNaseReveal™ Activity Assay Kits are fluorescent-based assays for the detection of RNase activity in liquid samples. Each kit utilizes a unique RNaseReveal™ Substrate, which is an RNA probe tagged with a fluorophore and a quencher, so that the intact probe is non-fluorescent. In the presence of RNase, the probe is cleaved and the fluorophore is detached from the quencher, releasing a fluorescent signal (Figure 1). The RNaseReveal™ Substrate in the original RNaseReveal™ Activity Assay Kit is tagged with a green fluorophore while that of the RNaseReveal™ Red Activity Assay Kit is tagged with a red fluorophore. The buffer and substrates have been formulated for maximum sensitivity, making the kits useful tools for quality control workflows or for detecting RNase contamination in solutions to be used with RNA work.

The original RNaseReveal[™] Activity Assay has a green fluorescence readout and is more sensitive than RNaseReveal[™] Red, with a lower limit of detection of ~0.08 pg RNase, using Biotium's RNase A (Cat. No. 99871-600uL) at ≥50 U/mg (Kunitz units). Full green substrate cleavage is achieved with ~800 pg of the same RNase A.

RNaseReveal™ Red features a unique RNase substrate with red fluorescence emission. It was designed for testing samples with intrinsic green fluorescence that interferes with a green fluorescent assay.

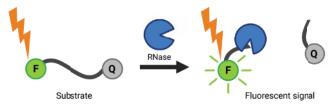


Figure 1. Mechanism of RNaseReveal™ Activity Assay Kits. The presence of RNase results in cleavage of the quencher from the substrate, causing a fluorescent signal. Created in https://BioRender.com

Spectral Properties

Kit	Cat. No.	Ex/Em (nm)	Fluorescence Readout
RNaseReveal™ Activity Assay Kit	31086	496/516	Green fluorescence
RNaseReveal™ Red Activity Assay Kit	31087	584/604	Red fluorescence

Experimental Protocol

Protocol outline

1. Incubate samples with substrate for 30 minutes at 37°C



2. Read initial fluorescence ("RFUinitial")



3. Add RNase and incubate 10 minutes at 37°C to maximally cleave the substrate



4. Read maximum fluorescence ("RFUmax")



5. Calculate the % of maximum (%max) signal for the initial fluorescence values

Materials required but not provided

- RNase-free water (see Related Products)
- RNase A (see Related Products)
- Black 96-well microplate

Assay protocol

When performing this assay for the first time, check to make sure your samples do not include any of the incompatible substances listed in Table 1 on page 2. It may also be helpful to include a well with 1 uL of RNase A added at the start of the assay in order to set an appropriate instrument gain (see notes to Step 5b).

- Allow RNaseReveal™ Substrate and 10X RNaseReveal™ Buffer to thaw and reach room temperature.
- Make a master mix of equal volumes of RNaseReveal™ Substrate and 10X RNaseReveal™ Buffer. Each well needs 20 uL of master mix (10 uL of Substrate + 10 uL of 10X Buffer).

Suggested experiment setup:

- a. Negative control (RNase-free water) two wells
- b. Test sample(s) two wells per test sample
- c. RNase-added control one well (optional for establishing microplate reader gain setting, see step 5 for details)

Note: If using this control, take special care to avoid cross-contamination of samples with RNase. Prepare the negative control and sample wells before preparing the RNase-added control.

- Pipette 20 uL of the master mix into the appropriate number of wells of a black 96-well plate.
- Pipette 80 uL of sample into each well. Mix by pipetting up and down. Pipette 80 uL of RNase-free water into 2 wells as a negative control. Pipette 1 uL of RNase A into one well, if you are doing the optional RNase-added control.
- Insert the plate into a fluorescent microplate reader and set up the RNaseReveal™ assay as follows:
 - a. Incubate for 30 minutes at 37°C (incubation can also be done externally in an incubator, protected from light).
 - b. Read fluorescence at Ex/Em 490/520 for the green assay or at Ex/Em 580/610 for the red assay. This value will be "RFUinitial".

Notes:

- i. Do not use autogain to maximize this signal. You will want to use the same setting later to read the "RFUmax" signal, which will be much brighter. Negative controls with RNase-free water are expected to have initial reads below 5% of the maximum signal.
- ii. The gain should be optimized for samples containing a saturating amount of RNase for detection of the fully cleaved substrate prior to running the experiment. When running the assay for the first time on a specific instrument, you may want to include a well with RNase added during the first incubation to set an appropriate gain to avoid saturation of the RFUmax reading.

- Add 1 uL RNase A (≥100 ng/mL) to each well. Mix by pipetting up and down.
- Return the plate to the microplate reader and resume the RNaseReveal™ program as follows:
 - a. Incubate for 5-10 minutes at 37°C (incubation can also be done externally in an incubator, protected from light).
 - b. Read fluorescence at Ex/Em 490/520 for the green assay or at Ex/Em 580/610 for the red assay. This value will be "RFUmax".
- Data analysis
 - a. Calculate the % of maximum signal (%max) for each sample's initial read using the following formula:

RFUinitial/RFUmax * 100

- b. A sample without RNase contamination will have a %max similar to the water control. The %max for the water control and other negative samples should be <5%. Samples with RNase contamination will have roughly 5-100% maximum signal.
- c. The RFUmax value for all samples should be similar to the water control. If the value is significantly lower, it may indicate that the sample contains a component that is inhibitory to the RNaseReveal™ assay. You may wish to prepare a dilution of your sample in RNase-free water and test the diluted sample in the assay.

Table 1. Incompatible Substances

Incompatible substances	Notes
Darkly colored solutions	Darkly colored solutions may interfere with excitation or emission of the RNaseReveal™ Substrate
Solutions that inhibit RNase activity	Solutions with high ionic strength (e.g., 5 M NaCl, 20X SSC, 3 M sodium acetate, etc.)
	Solutions with pH <4 or >9
	Chaotropic agents, detergents, chelating agents, or any solutions that denature proteins (e.g., SDS, guanidine thiocyanate, urea, EDTA, etc.)
Solutions that cause chemical instability of the RNaseReveal™ Substrate	Solutions with pH >9
	Caustic solutions (strong acids and bases, bleach)

Troubleshooting

Problem	Solutions	
Very low RFUmax values for samples	The RFUmax value for all samples should be similar to the water control. If the value is significantly lower, it may indicate that the sample contains a component that is inhibitory to the RNaseReveal™ assay. You may wish to prepare a dilution of your sample in RNase-free water and test the diluted sample in the assay.	
High %max values for water or negative controls		

Related Products

Cat. No.	Product
99871-600uL	RNase A
41024-4L	Water, Ultrapure Molecular Biology Grade
22028	RNase-X™ Decontamination Solution
31073	AccuBlue® Broad Range RNA Quantitation Kit
41044	EMBER™ Ultra RNA Gel Kit
41032	EMBER500™ RNA Prestain Loading Dye
28001	ExoBrite™ EV Total RNA Isolation Kit
CD506	CELLDATA RNAstorm™ 2.0 FFPE RNA Extraction Kit
CD510-96	CELLDATA RNAstorm™ 2.0 MagBead FFPE RNA Extraction Kit
CD504	CELLDATA RNAstorm™ Fresh Cell and Tissue RNA Isolation Kit
CD508	CELLDATA DNAstorm™/RNAstorm™ 2.0 Combination Kit

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